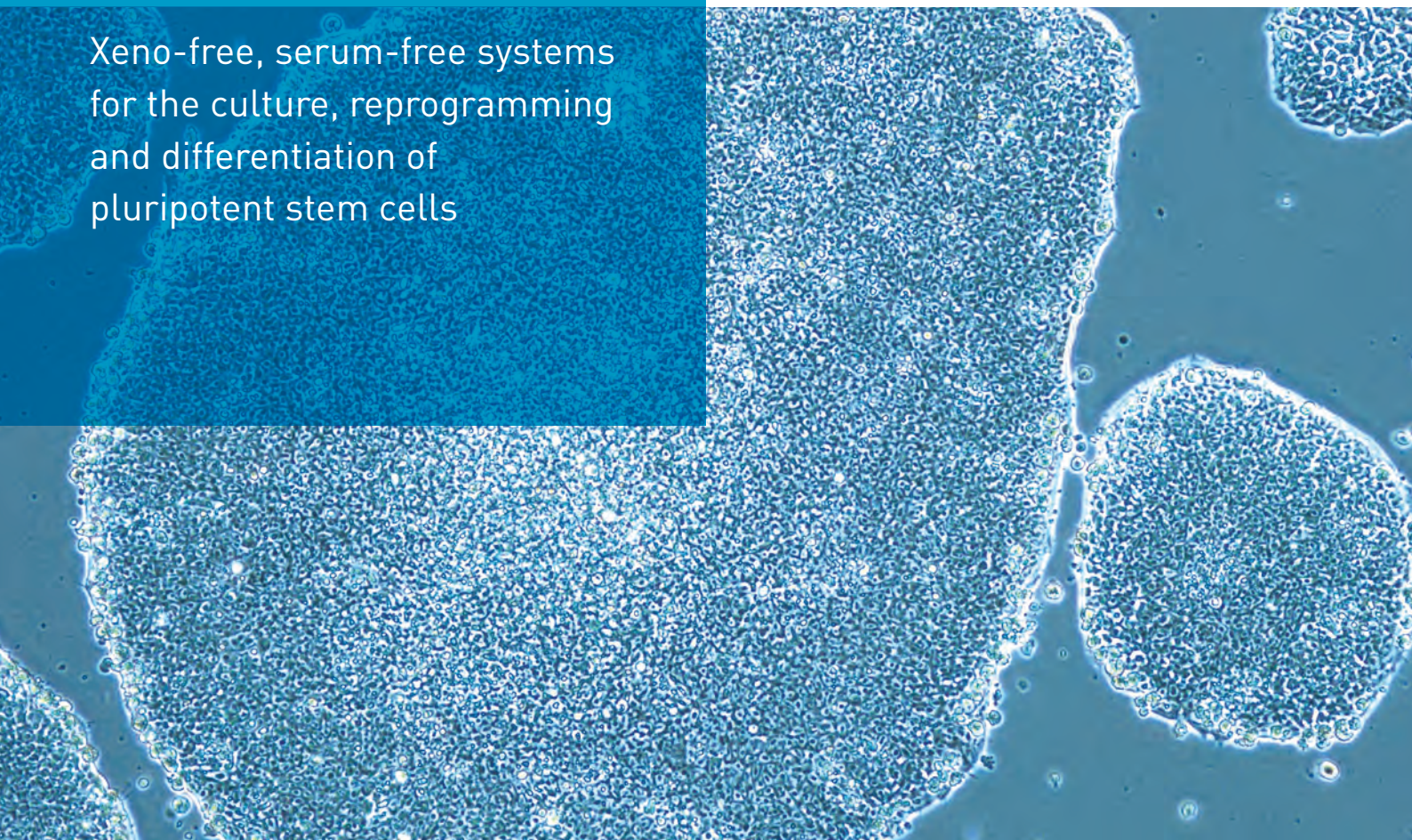


# Maintenance of Pluripotent Stem Cells

Xeno-free, serum-free systems  
for the culture, reprogramming  
and differentiation of  
pluripotent stem cells



# Choose the hPSC culture medium that's right for you

Human induced pluripotent cells (hiPSC) and human embryonic cells (hESC)

Feeder

Feeder-free

Which matrix do you use?

Laminin

Matrigel

Vitronectin

Single cell passaging

Enzyme-free passaging

NutriStem® hPSC XF culture medium



**NEW**  
NutriStem® V9 XF culture medium



100 µm

# From Research to Cell Based Therapies

The transition from stem cell culture research models to clinical applications requires the design and implementation of qualified processes. Defined, high-quality culture systems and appropriate documentation are therefore an essential element in the development of regenerative stem cell therapies, where implantation in humans is the desired outcome.

Biological Industries (BI) provides an optimized cell culture environment for human pluripotent stem cell research, including the NutriStem® defined, serum free (SF), xeno-free (XF) media family and its auxiliary reagents, manufactured in a cGMP compliant facility. In addition, a Drug Master File (DMF) registered at the FDA is available.

## Product Overview

### Media

#### NutriStem® hPSC XF

Defined, xeno-free, serum-free medium for optimal growth and expansion of hPSC on feeder or feeder-free conditions, using laminin or Matrigel.

#### NutriStem® V9 XF

Defined, xeno-free, NutriStem® V9 XF serum-free medium for optimal growth and expansion of hPSC on feeder-free conditions using vitronectin.

### Attachment

#### LaminStem™ 521

Defined, recombinant Laminin-521 for the attachment of human pluripotent stem cells in a feeder-free culture system.

#### Vitronectin ACF

Chemically defined, animal component-free (ACF) human recombinant lyophilized vitronectin protein for the attachment of human pluripotent stem cells in a feeder-free culture system.

### Dissociation

#### Recombinant Trypsin EDTA Solution

ACF recombinant trypsin solution with EDTA for efficient single cell dissociation of adherent cell types from surfaces and tissues.

#### EDTA Solution 0.5M

Enzyme-free, chemically defined, ACF dissociation solution.

### Cryopreservation

#### NutriFreez® D10 Cryopreservation Medium

ACF, protein-free and chemically defined freezing medium, for hPSC cryopreservation both as single cells and aggregates.

# hPSC Proliferation with NutriStem® hPSC XF

Product Name	Cat. No.	Size	Storage
NutriStem® hPSC XF	05-100-1A	500ml	-20°C
	05-100-1B	100ml	
NutriStem® hPSC XF Medium (Modified, GF-free, bFGF-free)	06-5100-01-1A	500ml	-20°C

Defined, xeno-free, serum-free medium designed to support the growth and expansion of hESC and hiPSC.

## Advantages

### Excellent performance

- Superior cell proliferation (low doubling time)
- Maintenance of pluripotent stem cell characteristics and stable karyotype over long term passages (>50 passages)

### User-friendly

- One bottle formulation
- Weekend-free feeding regime
- Straightforward adaptation protocol

### Flexible

- Versatile coating and culture methods
- Flexible packaging
- Custom modifications

### Defined, xeno-free, serum-free medium

- Reproducible and consistent results throughout experiments
- Batch-to-batch consistency

### cGMP medium

- Complete product dossier
- Registered DMF
- Produced under cGMP conditions

### Low growth factor concentrations (bFGF, TGF Beta)

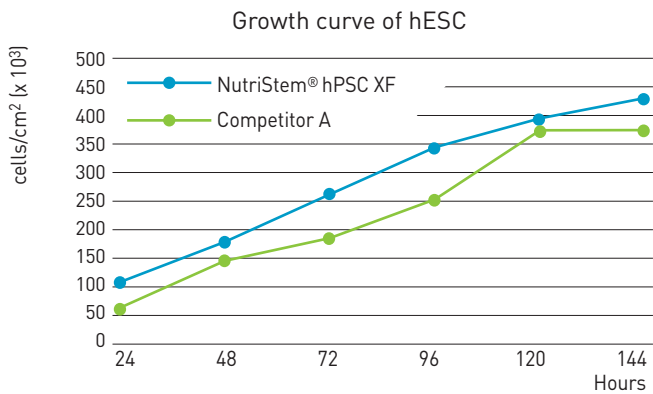
- Improves cell quality, reprogramming and differentiation capabilities

### Widely referenced in publications

- Feel confident in your research

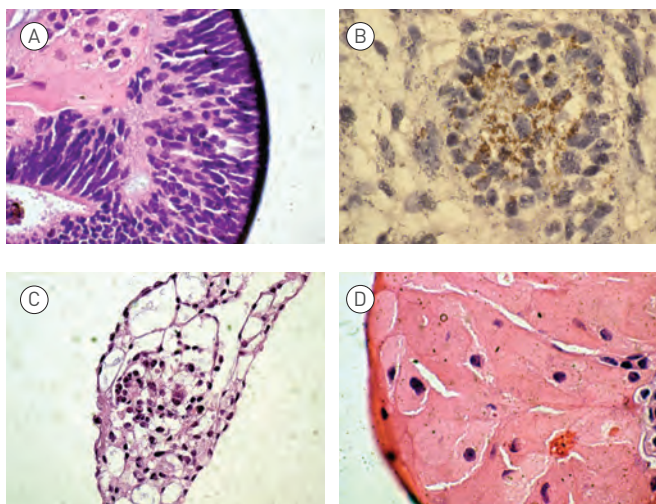
## Excellent proliferation of undifferentiated hPSC

NutriStem® hPSC XF enables excellent proliferation of undifferentiated hESC and hiPSC.



**Figure 1:** H1 cells (passage 6) were seeded in 96 well plates (Matrigel-coated) in the various media. Media were changed every 24 hours. The number of cells was determined using a CyQuant cell proliferation assay kit.

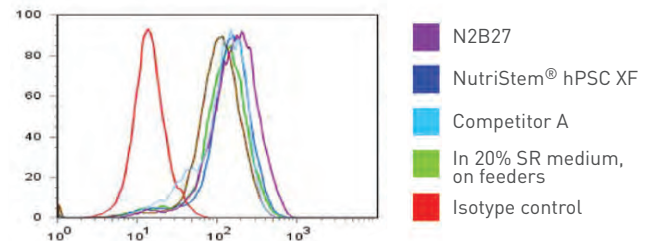
## Embryoid Body (EB) Formation



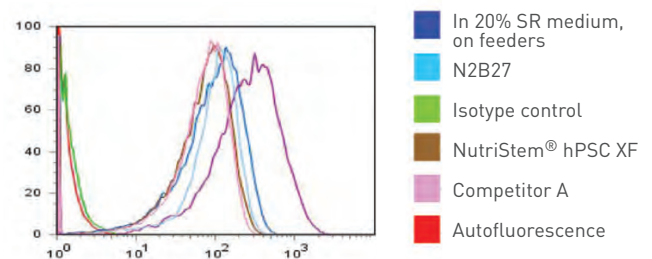
**Figure 2:** hESC from cell line H9.2 were cultured for 16 passages in NutriStem® hPSC XF using a Matrigel matrix and tested in vitro for pluripotency by EB formation. After suspension in serum supplemented medium the cells spontaneously formed embryoid bodies containing embryonic germ layers. Examining the histological sections of 14-day-old EBs, the following cell types were identified; (A) Neural rosette (ectoderm), (B) Neural rosette stained with Tubulin, (C) Primitive blood vessels (mesoderm) and (D) Megakaryocytes (mesoderm). Stained with H&E.

## High expression of pluripotent stem cell markers

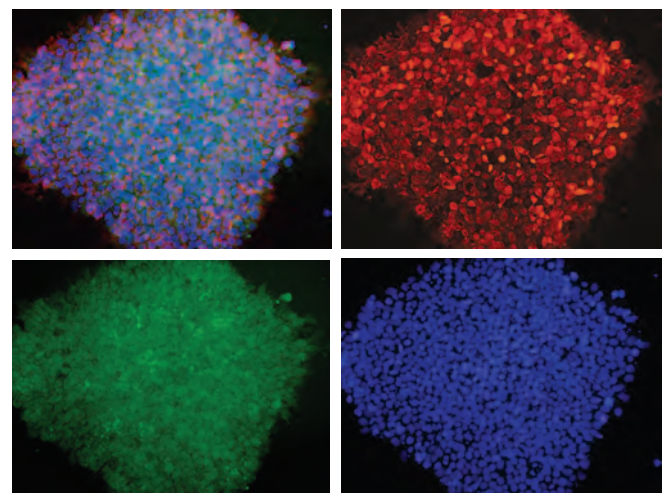
### Oct-4



### SSEA-4



**Figure 3:** H1 cells cultured in different media for 6 passages were analyzed and compared using flow cytometry and gene expression. Cells cultured in NutriStem® hPSC XF were found to be >90% positive for SSEA-4 and Oct-4.



**Figure 4:** H1 cell morphology and immunofluorescence analysis of hESC markers: red SSEA-4, green OCT4 and blue DAPI. H1 cells stained positive for the expression of pluripotency markers.

## NutriStem® hPSC XF gives you the freedom and versatility to derive and culture pluripotent stem cells in a variety of methods

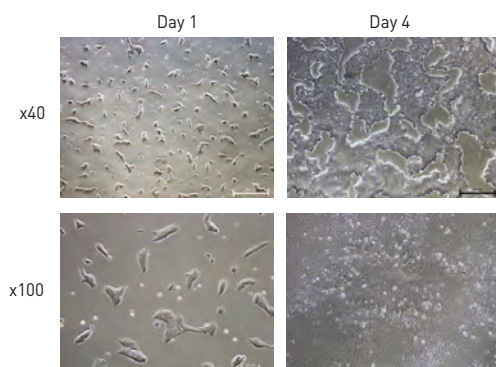
NutriStem® hPSC XF medium supports both feeder-dependent and feeder-free culture systems. The medium is also suitable for culture as colonies or monolayer, and supports single cell applications.

### Laminin-Based Culture System

LaminStem™ 521 with NutriStem® hPSC XF provide a superior culture environment for undifferentiated expansion and growth of hES and hiPS cells in a defined, xeno-free, and feeder-free culture system as a monolayer, while maintaining proper phenotype and genetic stability. Studies have shown that efficient clonal derivation of hES cell lines is possible with the combined use of NutriStem® hPSC XF medium and LaminStem™ 521 substrate, finding that the cells grew better in NutriStem® hPSC XF than any other defined medium tested, and that hES cells can be passaged and maintained using a single-cell expansion protocol (Rodin, S. et al. 2014).

### Single cell passaging using LaminStem™ 521 and Recombinant Trypsin EDTA Solution

Culturing of hPSC using NutriStem® hPSC with LaminStem™ 521 enables easy and reliable single-cell passaging without artificial apoptosis inhibitors, such as ROCK inhibitor (Y-27632). This provides standardized procedures that are fast and easy to use. For the efficient dissociation and passaging Recombinant Trypsin EDTA Solution should be used.



**Figure 5:** Typical recovery of H1 (61) hESC from single-cell passage using Recombinant Trypsin EDTA Solution and NutriStem® hPSC XF medium on 0.5µg/cm<sup>2</sup> LaminStem™ 521. Representative images for colony morphology one day and 4 days post-passage.

“hES cells grew better in the xeno-free chemically defined NutriStem® hESC XF”

(Rodin et al. 2014)

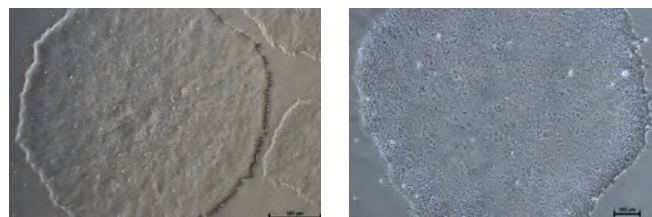
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- Rodin S, et al. Clonal culturing of human embryonic stem cells on laminin-521/E-cadherin matrix in defined and xeno-free environment. *Nat Commun.* 5:3195. doi: 10.1038/ncomms4195, 2014

### Clinical Applications

- Hovatta, Outi. Infectious problems associated with transplantation of cells differentiated from pluripotent stem cells. *Seminars in Immunopathology: Volume 33, Issue 6, pp 627-30, April 2011*

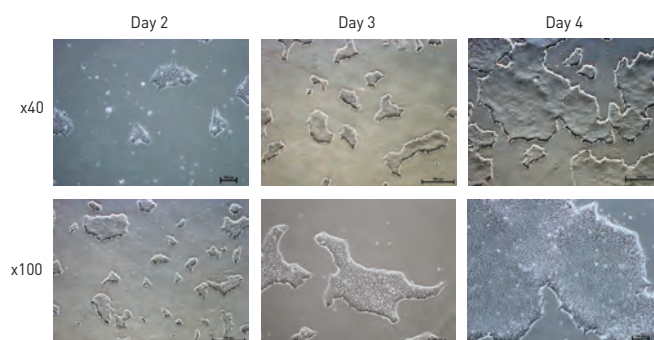
### Matrigel™-Based Culture System



**Figure 6:** H1 hESC cultured in NutriStem® hPSC XF on Matrigel™ display compact colonies and distinct colony morphology typical of hPSC.

### Enzyme-free passaging with EDTA

Small aggregate dissociation using EDTA is a gentle, enzyme-free method of passaging cells grown in feeder-free conditions.



**Figure 7:** Typical recovery of hESC from enzyme-free passage (0.5mM EDTA) using NutriStem® hPSC XF medium on Matrigel™. Representative results for colony morphology of H1 hESC 2-4 days post-passage.

## Key References

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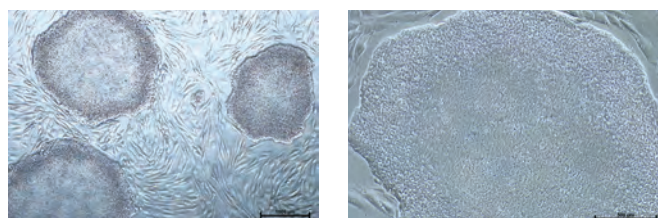
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- J. Lenzi et al. ALS mutant FUS proteins are recruited into stress granules in induced Pluripotent Stem Cells (iPSCs) derived motoneurons. *Disease Models & Mechanisms*: 8, 755-766, 2015

### Clinical Applications

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## Feeder-Dependent Culture (MEF/HFF)



**Figure 8:** H1 hESC colonies on MEF feeder layer display compact colonies and distinct colony morphology typical of hPSC.

“NutriStem appears to support iPSC culture on feeders better than E8”

(T. Cerbini et al., 2015)

## Key References

### Gene Editing

- T. Cerbini et al., Transfection, Selection, and Colony-picking of Human Induced Pluripotent Stem Cells TALEN-targeted with a GFP Gene into the AAVS1 Safe Harbor, *JoVE (Journal of Visualized Experiments)*, 2015

### Clinical Applications

- P. Menasché et al. Human embryonic stem cell-derived cardiac progenitors for severe heart failure treatment: first clinical case report. *European heart journal* (2015): ehv189
- Y. Luo et al., Stable Enhanced Green Fluorescent Protein Expression After Differentiation and Transplantation of Reporter Human Induced Pluripotent Stem Cells Generated by AAVS1 Transcription Activator-Like Effector Nucleases. *STEM CELLS Translational Medicine: Volume 3, Issue 7*, pp 821-35, 2014

## Auxiliary Products

Product Name	Cat. No.	Size	Storage
EDTA Solution 0.5M	01-862-1B	100ml	RT

Diluted EDTA Solution 0.5mM is an enzyme-free, chemically defined, Animal Component Free (ACF) solution, suitable for the dissociation of human pluripotent stem cells. EDTA Solution 0.5mM mediates rapid cell dissociation by chelating calcium and magnesium ions that facilitate cell adhesion

Product Name	Cat. No.	Size	Storage
LaminStem™ 521	05-753-1F	1ml	-20°C

LaminStem™ 521 facilitates self-renewal hPSC in a defined, feeder-free and xeno-free cell culture system. LaminStem™ 521 is composed of purified laminin-521, a cell-type specific basement membrane protein proven to support excellent attachment proliferation of hES and hiPS cells.

Product Name	Cat. No.	Size	Storage
Recombinant Trypsin	03-079-1B	100ml	RT
EDTA Solution	03-079-1C	20ml	

Recombinant Trypsin EDTA Solution was developed for efficient single cell dissociation of adherent cell types from surfaces and tissues and were optimized for sensitive cells, such as hPSC.

Recombinant Trypsin EDTA Solution is ready-to-use and animal component free. The addition of EDTA accelerates the dissociation phase. The solution does not contain any chymotrypsin, carboxypeptidase A, or other protease contaminants.

# hPSC Proliferation with NutriStem<sup>®</sup> V9 XF

<b>Product Name</b>	<b>Cat. No.</b>	<b>Size</b>	<b>Storage</b>
NutriStem <sup>®</sup> V9 XF basal medium	05-105-1A	500ml	-10° to -20°C
NutriStem <sup>®</sup> V9 XF Supplement Mix	05-106-1F	1ml	-10° to -20°C

Defined, xeno-free, serum-free culture medium for hPSC optimized for vitronectin.

## Advantages

### Excellent performance

- Superior proliferation rates in long-term culture on vitronectin coated culture ware
- Maintenance of pluripotent stem cell characteristic over long term passages (>30 passages)
- Supportive for difficult lines and routine culture

### User-friendly

- Weekend-free feeding regime
- Pre-coating free protocol with BI's ACF Vitronectin
- ROCK inhibitor free protocol for seeding, passaging and thawing

### Defined, xeno-free, serum-free medium

- Reproducible and consistent results throughout experiments
- Batch-to-batch consistency

### cGMP media

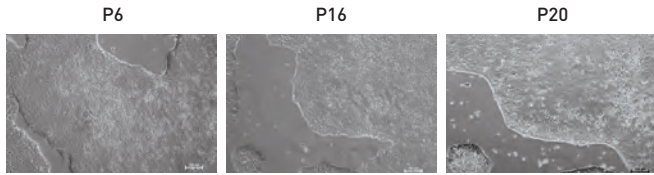
- Produced under cGMP conditions

### Cytokine-free basal medium

- Improves cell quality and ability to reprogram and differentiate

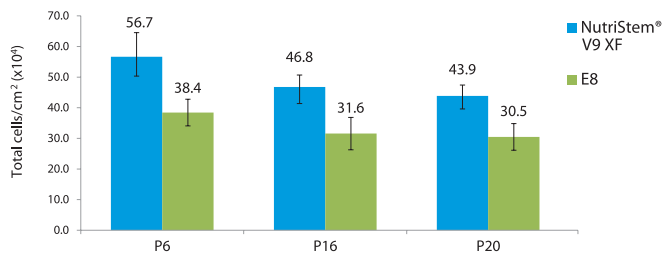


## Typical cell morphology during long-term culture



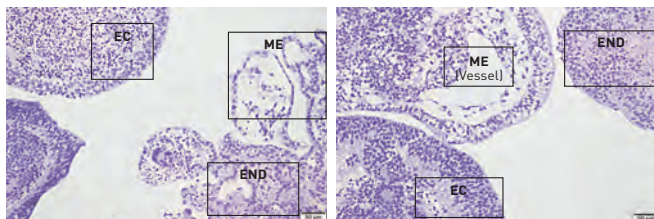
**Figure 9:** Phase contrast images (x100) of H1 hESC culture maintained in NutriStem® V9 XF using 0.5µg/cm<sup>2</sup> Vitronectin ACF-coated cultureware, show typical morphology.

## Superior proliferation rate in long-term culture



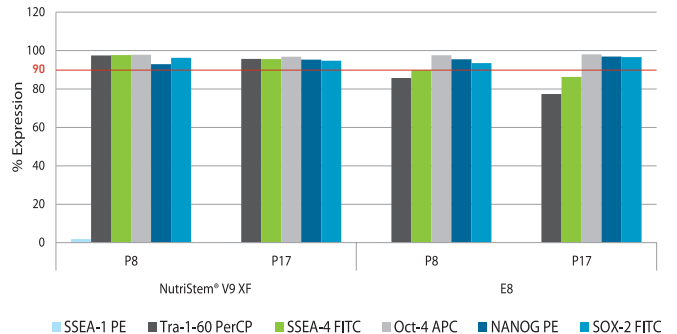
**Figure 10:** Nucleocounts performed on hESC suspension (Chemometec) during long-term expansion in NutriStem® V9 XF compared to E8 using 0.5µg/cm<sup>2</sup> Vitronectin ACF and passage as small aggregates every 3-5 days using 0.5mM EDTA solution.

## Embryoid body (EB) Formation

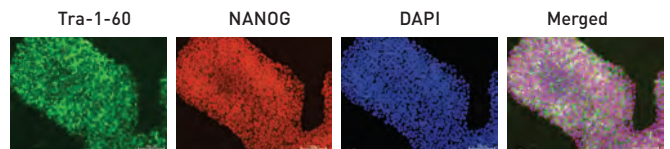


**Figure 11:** Embryoid bodies (EBs) generated from H1 hESC expanded for 6 passages in NutriStem® V9 XF medium on Vitronectin ACF as an evaluation of pluripotency. Cells were suspended in NutriStem® V9 XF basal, where they spontaneously formed EBs, for 18 days. Cell types identified by examination of EBs histological sections stained with H&E. See letters EC=neural rosettes, ME=primitive vessels, ED=primitive parenchyma (X100)

## High expression of pluripotent stem cell markers



**Figure 12:** Immunophenotyping analysis of human pluripotent markers of H1 hESC cultured in NutriStem® V9 XF at P8 and P17 shows elevated marker expression over 90%. Data presented as % expression from gated viable cells.



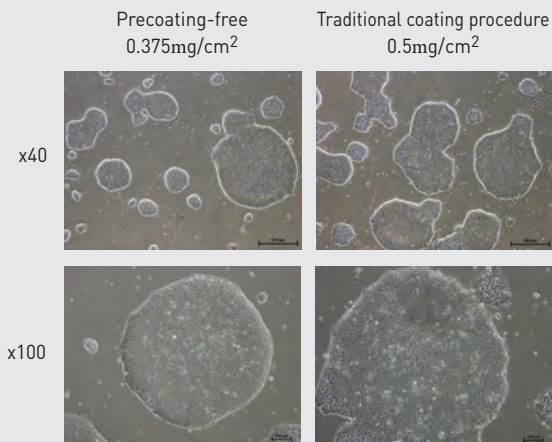
**Figure 13:** Immunofluorescence analysis of human pluripotent markers of H1 hPSC expanded in NutriStem® V9 XF medium under a weekend-free feeding regime at P8. Cells were stained for pluripotent surface markers: TRA 1-60 (Alexa Fluor) (green) and nuclear conjugated markers: NANOG-RRX (red), counterstained with DAPI (blue). Scale bar 200µm.

## Save time with precoating-free procedure!

A user-friendly protocol has been developed to eliminate the precoating procedure. While seeding, vitronectin is added directly into NutriStem® V9 XF medium, making precoating unnecessary.

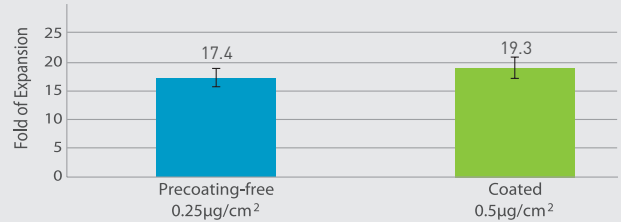
Novel precoating-free procedure		Traditional coating procedure	
Actual Vitronectin ACF coating	Vitronectin ACF volume added to 3ml medium	Vitronectin ACF coating	Vitronectin ACF volume
0.25-0.375µg/cm <sup>2</sup>	5-7.5µl	0.5µg/cm <sup>2</sup>	10µl
~30 minutes		90 minutes minimum	

## Typical cell morphology



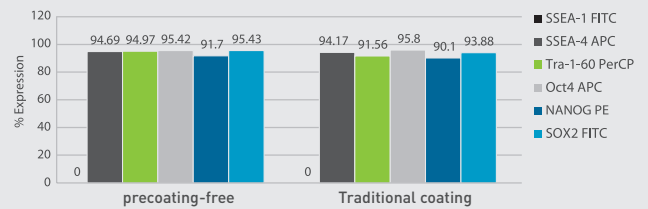
**Figure 14:** Colony morphology of hPSC cultured in NutriStem® V9 XF for 11 sequential passages using coated and precoating-free protocol (Vitronectin ACF added directly to NutriStem® V9 XF culture medium before cells seeding).

## High proliferation rate of hPSC during long-term expansion

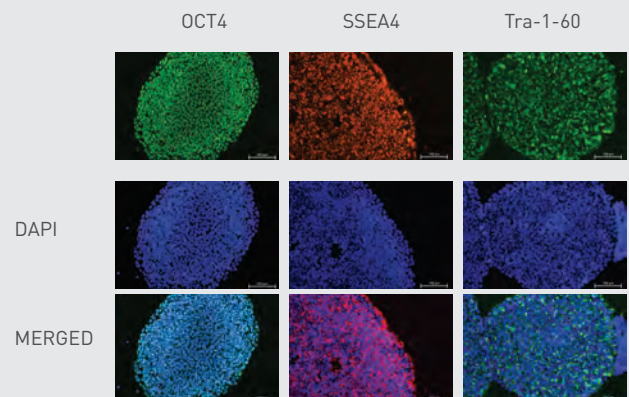


**Figure 15:** Nucleocounts performed on cells suspension (Chemometec) in NutriStem® V9 XF using traditional coating and precoating-free protocol and passaged as aggregates using 0.5mM EDTA solution. Equivalent proliferation rates were measured in both methods.

## High expression of pluripotent markers



**Figure 16:** Flow cytometry analysis of H1 cultured in NutriStem® V9 XF for 11 passages using precoating-free protocol. Vitronectin ACF was added directly to NutriStem® V9 XF culture medium before cells seeding. High expression of pluripotent markers, over 90% is measured. Data presented as % expression from gated viable cells.



**Figure 17:** Immunofluorescence analysis of human pluripotent markers of H1 hPSC expanded in NutriStem® V9 XF medium using the precoating-free protocol for 8 passages. Cells from P8 were fixed and stained for the classical pluripotent surface markers: SSEA4 (RRX) (red), TRA-1-60 (Alexa fluor) (green) and nuclear conjugated markers: OCT-4-Alexa fluor, counterstained with DAPI (blue). Scale bar 200 µm.

## Auxiliary Products

Product Name	Cat. No.	Size	Storage
Vitronectin ACF	05-754-0002	200mg	-20° to -80°C

Vitronectin is a secreted glycoprotein that supports cell adhesion through binding to various integrins and proteoglycans. Vitronectin ACF (Animal Component Free) can function as a chemically-defined matrix component for the attachment of human embryonic and induced pluripotent stem cells in a feeder-free culture system. Vitronectin ACF is a 459 amino acid, single-chain, monomeric recombinant protein, which migrates at an apparent molecular weight of 75 kDa by SDS-PAGE under reducing conditions. The calculated molecular weight of Vitronectin ACF is 52.2 kDa.

Product Name	Cat. No.	Size	Storage
0.5M EDTA Solution	01-861-1B	100ml	RT

Diluted 0.5mM EDTA solution is an enzyme-free, chemically defined, ACF solution, Diluted 0.5mM EDTA solution suitable for the dissociation of human pluripotent stem cells. Diluted 0.5mM EDTA solution rapid cell dissociation by chelating calcium and Diluted 0.5mM EDTA solution magnesium ions that facilitate cell adhesion.



# hPSC Cryopreservation

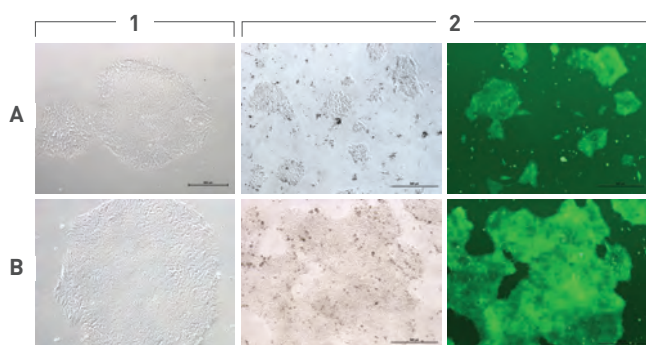
Product Name	Cat. No.	Unit Size	Storage
NutriFreez® D10 Cryopreservation Medium	05-713-1A	500ml	2-8°C
	05-713-1B	100ml	
	05-713-1C	20ml	
	05-713-1D	10ml	
	05-713-1E	50ml	

NutriFreez® D10 Cryopreservation Medium is an animal components-free, ready-to-use solution for the cryopreservation of animal cells.

NutriFreez® D10 Cryopreservation Medium was developed to maintain ACF conditions during cryopreservation when culturing cells in a XF culture system, and has been extensively validated with human ES cells (H1, H9 and HuES9). NutriFreez® D10 Cryopreservation Medium has shown to be very effective for the cryopreservation of hPSCs as single cells and cell aggregates. Cells preserved with NutriFreez® D10 Cryopreservation Medium show high viability, attachment, growth performance, and maintenance of pluripotency markers after thawing (Figure 1), with superior results compared to both serum-containing freezing media and other serum-free solutions.

## Advantages

- Chemically defined, Animal component-free (ACF), Protein-free
- Works with various media
- Suitable for freezing hESC and hiPSC cultured in both feeder and feeder-free conditions
- High recovery efficiency: maintains excellent attachment ability as well as growth performance
- Maintains hESC and hiPSC pluripotency
- Complete formulation; Ready-to-use at 2-8°C
- For cryopreservation of hPSC clumps or single cells



**Figure 18:** H1 hES cells (1) and BG01V/hOG (2) GFP reporter cells frozen in NutriFreez® D10 Cryopreservation Medium. Cryopreserved hES cells were thawed into NutriStem® hPSC Medium on Matrigel-coated plates. Cells show high viability at day 1 (A) and at day 4 post-thaw (B).

“...cryopreservation with CryoStem\* showed the best recovery rate for hPSCs after thawing”

(Nishishita N, et al., 2015)

\* NutriFreez® replaces the brandname “CryoStem”

## Key References

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- N. Nishishita et al. Generation of Virus-Free Induced Pluripotent Stem Cell Clones on a Synthetic Matrix via a Single Cell Subcloning in the Naïve State. PLoS ONE 7(6): e38389. doi:10.1371/journal.pone.0038389, 2012
- F. Pistollato et al. Standardization of pluripotent stem cell cultures for toxicity testing. Vol. 8, No. 2, Pages 239-257 (doi:10.1517/17425255.2012.639763), 2012

# Key References for Derivation, Reprogramming and Differentiation

## hESC Derivation

NutriStem® hPSC XF medium enables successful derivation of new hESC lines, as well as long-term genetically stable growth of the clonal hESC lines in chemically defined, xeno-free environment.

### Key References

- M.V. Krivega et al. Cyclin E1 plays a key role in balancing between totipotency and differentiation in human embryonic cells. *MHR: Basic science of reproductive medicine*, Volume 21, Issue 12, 1 December 2015
- S. Rodin et al., Monolayer culturing and cloning of human pluripotent stem cells on laminin-521-based matrices under xeno-free and chemically defined conditions. *Nature Protocols* 9, 2354–2368 (2014) doi:10.1038/nprot.2014.159

## hiPSC Reprogramming

mRNA-based cellular reprogramming of human cells  
NutriStem® hPSC XF medium supports mRNA-based cellular reprogramming of human cells. mRNA reprogramming is a fast, safe and efficient method for generating integration-free, virus-free, clinically relevant iPSC cell lines from mature human cells. BI also offers the possibility for modified NutriStem® hPSC XF without growth factors.  
Clonal mRNA reprogrammed iPSC lines can be expanded and maintained in NutriStem® hPSC XF.

### Key References

- Protocol describes using laminin substrate and NutriStem™ hPSC XF Culture Medium to provide a complete xeno-free reprogramming environment:  
Protocol: Stemgent® StemRNA™-NM Reprogramming Kit for Reprogramming Adult and Neonatal Human Fibroblasts, ReproCell
- iPSC generation by reprogramming EPCs using self-replicative RNA (srRNA):  
X. Gao et. al. Comparative transcriptomic analysis of endothelial progenitor cells derived from umbilical cord blood and adult peripheral blood: Implications for the generation of induced pluripotent stem cells. *Stem Cell Research*, 2017
- Efficient Reprogramming of Human Fibroblasts and Blood-Derived Endothelial Progenitor Cells:  
Poleganov M. A. et. al. *Human Gene Therapy*. August 2015, 26(11): 751-766. <https://doi.org/10.1089/hum.2015.045>

- L. Healy, L Ruban, Derivation of Induced Pluripotent Stem Cells, *Atlas of Human Pluripotent Stem Cells in Culture*, pp 149-165. Springer US 2015
- IVT-RNA-based reprogramming method:  
S. Herz, Optimization of RNA-based transgene expression by targeting Protein Kinase R. Dissertation for the degree "Doctor rerum naturalium", 2015
- Generation of stable, pluripotent ESC-iPS and fibroblast-iPS cell lines:  
S. Eminli-Meissner et al. A novel four transfection protocol for deriving iPS cell lines from human blood- derived endothelial progenitor cells (EPCs) and adult human dermal fibroblasts using a cocktail of non-modified reprogramming and immune evasion mRNAs. Scientific Poster, REPROCELL, 2015
- M. Brouwer et al. Choices for Induction of Pluripotency: Recent Developments in Human Induced Pluripotent Stem Cell Reprogramming Strategies. *Stem Cell Reviews and Reports: Volume 12, Issue 1*, pp 54–72, 2015
- Reprogramming keratinocytes to pluripotency:  
L. Warren et al., Highly Efficient Reprogramming to Pluripotency and Directed Differentiation of Human Cells with Synthetic Modified mRNA. *Cell Stem Cell* 7 (5): 618–630 (2010)
- Reprogramming of human and mouse adipose-derived stem cells into iPSC:  
S. Sugii et al., Human and mouse adipose-derived cells support feeder-independent induction of pluripotent stem cells. *PNAS* February 23, 2010 vol. 107 no. 8 3558-3563

## hPSC Differentiation

NutriStem® hPSC XF is widely referenced in publications, showing effective differentiation of hPSC into variety of cell types.

### Key References

#### Cardiomyocyte differentiation

- R. Ophir et al. Inflammation And Contractility Are Altered By Obstructive Sleep Apnea Children's Serum, In *Human Embryonic Stem Cell Derived Cardiomyocytes*. *American Journal of Respiratory and Critical Care Medicine* 2017
- J. KRISTENSSON, Optimization of Growth Conditions for Expansion of Cardiac Stem Cells Resident in the Adult Human Heart. Master's thesis in Biotechnology, Department of Physics, Division of Biological Physics, Chalmers University of Technology, Gothenburg, Sweden 2016
- S. Rajasingh et al. Generation of Functional Cardiomyocytes from Efficiently Generated Human iPSCs and a Novel Method of Measuring Contractility. *PLoS one* 10.8, 2015: e0134093 (Fibroblast origin)
- L. Jacquet et al. Three Huntington's Disease Specific Mutation-Carrying Human Embryonic Stem Cell Lines Have Stable Number of CAG Repeats upon In Vitro Differentiation into Cardiomyocytes. *PLoS one* 10.5, 2015 V. Bellamy et al., Long-term functional benefits of human embryonic stem cell-derived cardiac progenitors embedded into a fibrin scaffold, *The Journal of Heart and Lung Transplantation*,

2014

- E. Di Pasquale et al. Generation of human cardiomyocytes: a differentiation protocol from feeder-free human-induced pluripotent stem cells. *JoVE (Journal of Visualized Experiments)* 76 (2013): e50429–e50429
- G. Földes and M. Mioulane. High-content imaging and analysis of pluripotent stem cell-derived cardiomyocytes. *Imaging and Tracking Stem Cells*. Humana Press, 2013.
- P.W. Burridge and E.T. Zambidis. Highly efficient directed differentiation of human induced pluripotent stem cells into cardiomyocytes. *Pluripotent Stem Cells: Methods and Protocols*. Methods in Molecular Biology, volume 997, pp 149–161, Humana Press, 2013.

#### Neuronal differentiation

- K.M. Gray et al. Self-oligomerization regulates stability of Survival Motor Neuron (SMN) protein isoforms by sequestering an SCFS<sup>lmb</sup> degron. *Molecular Biology of the Cell*, 2017 mbc.E17-11-0627
- X. Yuan et al. A hypomorphic PIGA gene mutation causes severe defects in neuron development and susceptibility to complement-mediated toxicity in a human iPSC model, *PLOS ONE*, 2017
- R. De-Santis, A. Rosa et al. FUS Mutant Human Motoneurons Display Altered Transcriptome and microRNA Pathways with Implications for ALS Pathogenesis. *Stem Cell Reports* (2017), <https://doi.org/10.1016/j.stemcr.2017.09.004>
- D. VOULGARIS, Evaluation of Small Molecules for Neuroectoderm differentiation & patterning using Factorial Experimental Design. Master Thesis in Applied Physics, Department of Physics, Division of Biological Physics, Chalmers University of Technology, Göteborg, Sweden 2016
- P. Bergström et al. Amyloid precursor protein expression and processing are differentially regulated during cortical neuron differentiation, *Scientific Reports*, 2016
- Tieng, V. et al. Elimination of proliferating cells from CNS grafts using a Ki67 promoter-driven thymidine kinase, *Molecular Therapy — Methods & Clinical Development* 6, Article number: 16069, 2016
- Brykczynska, U, et al. CGG Repeat-Induced FMR1 Silencing Depends on the Expansion Size in Human iPSCs and Neurons Carrying Unmethylated Full Mutations, *Stem Cell Reports*, 2016
- Sellgren, C.M. et al. Patient-specific models of microglia-mediated engulfment of synapses and neural progenitors *Molecular Psychiatry*, 2016
- K. Alessandri et al. A 3D printed microfluidic device for production of functionalized hydrogel microcapsules for culture and differentiation of human Neuronal Stem Cells (hNSC). *Lab on a Chip*: 16(9), 2016
- Cosset, E. et al. Human tissue engineering allows the identification of active miRNA regulators of glioblastoma aggressiveness, *Biomaterials*, 2016
- M. Di Salvio et al. Pur- $\alpha$  functionally interacts with FUS carrying ALS-associated mutations. *Cell Death & Disease*, 2015
- A. J. Schwab, A.D. Ebert, Sensory Neurons Do Not Induce Motor Neuron Loss in a Human Stem Cell Model of Spinal Muscular Atrophy. *PLoS One*. 2014; 9(7): e103112
- H.X. Nguyen et al., Induction of early neural precursors and derivation of tripotent neural stem cells from human pluripotent stem cells under xeno-free conditions. *Journal of Comparative Neurology*:

Volume 522, Issue 12, pp 2767–2783, 2014

- Lenzi, J., et al. Differentiation of control and ALS mutant human iPSCs into functional skeletal muscle cells, a tool for the study of neuromuscular diseases. *Stem Cell Research: Volume 17, Issue 1, Pages 140–147*, 2016.

#### Retinal differentiation

- R.A. Hazim et al. Differentiation of RPE cells from integration-free iPSC cells and their cell biological characterization. *Stem Cell Research & Therapy* 2017
- E. Welby et al. Isolation and Comparative Transcriptome Analysis of Human Fetal and iPSC-Derived Cone Photoreceptor Cells. *Stem Cell Reports* (2017), <https://doi.org/10.1016/j.stemcr.2017.10.018>
- S. Petrus-Reurer et al. Integration of Subretinal Suspension Transplants of Human Embryonic Stem Cell-Derived Retinal Pigment Epithelial Cells in a Large-Eyed Model of Geographic Atrophy. *Retinal Cell Biology*, February 2017
- A. Reyes, et al. Xeno-Free and Defined Human Embryonic Stem Cell-Derived Retinal Pigment Epithelial Cells Functionally Integrate in a Large-Eyed Preclinical Model Plaza. *Stem Cell Reports: Volume 6, Issue 1, p9–17*, 2015

## Ordering information

Product Name	Cat. No.	Size	Storage
NutriStem® hPSC XF	05-100-1A	500ml	-20°C
	05-100-1B	100ml	
NutriStem® hPSC XF Medium (Modified, GF-free, bFGF-free)	06-5100-01-1A	500ml	-20°C
NutriStem® V9 XF Basal Medium NutriStem® V9 XF Supplement Mix	05-105-1A	500ml	-20°C
	05-106-1F	1ml	
LaminStem™ 521	05-753-1F	1ml	-20°C
Vitronectin ACF	05-754-0002	1ml	-20° to -80°C
Recombinant Trypsin Solution	03-078-1A	500ml	RT
	03-078-1B	100ml	
Recombinant Trypsin EDTA Solution	03-079-1A	500ml	RT
	03-079-1B	100ml	
EDTA Solution 0.5M	01-862-1B	100ml	RT
Accutase Solution	03-073-1B	100ml	-20°C
NutriFreez® D10 Cryopreservation Medium	05-713-1A	500ml	2-8°C
	05-713-1B	100ml	
	05-713-1C	20ml	
	05-713-1D	10ml	
	05-713-1E	50ml	



Biological Industries (BI) has been committed for over 30 years to provide optimal and innovative solutions for cell culture practice.

BI manufactures and supplies life science products to biopharmaceutical, academic and government research facilities, as well as to biopharma companies.

**Our diverse portfolio of products and services includes all of the following:**

- Liquid and powdered cell culture media
- Sterile sera (foetal bovine serum, newborn calf serum, donor horse, etc.)
- Novel serum-free and animal component-free media and supplements
- Products for stem cell research and cell-based therapies
- Products for cytogenetics
- Products for mycoplasma detection and treatment
- Disinfectants
- Products for molecular biology
- custom formulations and contract manufacturing services

All BI's products are manufactured via a quality management system ISO 9001:2015 and in regards of medical devices ISO 13485:2016. All aspects of the products life cycle fall under the QMS procedures. The set-up of clean zone and clean room facilities for manufacturing are following ISO 14644, whereas the production rooms are ISO 8, storage of sterile accessories ISO 7 and filling rooms ISO 5. Aseptic filling and validation is performed according to ISO 13408.

BI exports its products to more than 50 countries worldwide, via a network of exclusive distributors. Over the years we have established a reputation for fast delivery, and excellent technical support.

From the outset, the policy of BI has been based on the need to maintain an active Research and Development program in all facets of company activities. The company has its own in-house R&D department, and in addition, maintains active contact with science-based companies and research institutions in Israel and abroad, including know-how agreements with several such institutions. These ongoing efforts have led to the introduction of a series of serum-free medium products, as well as many other products for cell culture and molecular biology.

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